

UC Extension Mol Biol X107A Test #2, Due 11/16/00.

Name:
(Please print)

Total points: 18

Take home test, **Ch 6-11** (consistent with class coverage of these chapters, as presented in the handouts, and noted at the web site Updates page).

A sketch similar to Fig 7.11 is included; you should understand the general role of the regulatory proteins.

DUE: Thurs 11/16, at start of class. For “emergency use only”... You may return the test by mail (postmarked by the due date): <address>. Do NOT send by any method that requires my signature to pick it up!

The general idea is that a take home test is just like a test in class, except that you determine the time and place to take it.

Do not open the test until you are ready to start taking it. Opening the test marks the end of your study time and the beginning of test time. Prior to beginning the test, you may study as much as you wish, and in any way. You may study with others, so long as no one involved has yet opened the test. You may not discuss the test with others (except me) until after the due time.

Test is closed book. **You may use one page of notes of your own composition; no direct copies of book material allowed (see web site for more information about the note page).** You may supply extra paper. Otherwise, you may NOT use books, handouts, personal notes, the string, or other outside sources or props.

This test is similar to other tests in this course (including the sample). Try to resolve doubts about the meaning of test questions by considering them in the context of this course. If you feel there are ambiguities in a question, state any assumptions you make.

Estimated time: 1-2 hour. It is not a timed test. Take the time you need to do it well. As long as you feel you are being productive, you are welcome to continue. (That also means you need to know when to stop.) I would appreciate it if you would note how long you took.

You should take the test “straight through” in one session. However, you may seek clarification of test questions at a reasonable time after your primary session.

Feel free to call me at home, either in preparation for the test or if you need clarification of questions during the test. I will “guarantee” phone availability 7-9 pm Wednesday evening and 11-12 Thursday morning. If all else fails, come a few minutes early Thursday evening, and ask me remaining questions before class.

⇒ Please SIGN the following agreement (often called an Honor System pledge):

I agree to take this test according to the rules established for it.
(-signature)

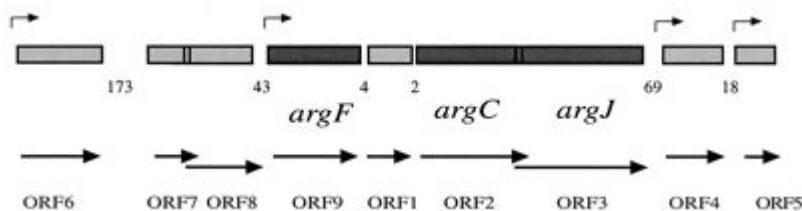
Total points: 18

General

- Closed book. You may use one page of your own notes.
- Your reasons are as important as your answers. If in doubt, say more, not less, about what you do and why.
- If you change your mind, please make clear what I'm supposed to read. If you use extra paper, put your name and the question number on it, and refer to the extra page on the test sheet.

1. (1 pt.) How is it that transcription can induce positive supercoils (overwinding) in part of the template? Which part? (Ignore other factors, such as topoisomerases.)

2. (6 pts.) (continues on next page) The following Fig is from a recent paper. It is a map of several contiguous genes in a bacterium. The genes are simply named ORF with a number; ORF means "open reading frame", a name for a region that appears to be a gene, but has not yet been specifically identified. Some of the ORFs were later shown to be part of the pathway for synthesizing the required amino acid nutrient, arginine; these genes are also labeled with the name *arg*. The bent arrows at the top identify transcriptional start sites.



The numbers below the top map (173, etc) are the number of nucleotides between genes. You can mostly ignore these, though one will be somewhat useful in part a.

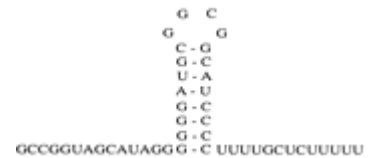
All parts of this question are intended to be about transcription and its regulation, from chapters officially covered by this test. They are not intended to be about translational issues. (The article itself is about such issues, in part, and you may think they are relevant. But it would be unfair for the test to grade on any translational issues here.)

⇒ The parts of this question (next page) are mostly independent.

a. (2 pts.) They show that a regulatory protein binds a site overlapping the argF promoter. Part of their evidence was a footprint experiment, which shows protection of a 29 base region, which extends from the middle of the -10 region to somewhat upstream of the -35 region. Sketch the result of the footprint experiment that would lead to this conclusion. Be sure to include a control. Label key features of your sketch well enough to make the main point. Relate the sketch to the map shown above. (That is, show what region of the top map is involved in this part.)

b. (1 pt.; no credit without explanation) Judging from the information given in part a, is it most likely that this protein is a positive control protein or a negative control protein? Explain; that is, how can you tell from the given information? Be sure your explanation is clear enough to distinguish what the two terms mean.

c. (2 pts.) The Figure at the right shows part of an RNA from a region near the beginning of ORF5. What is the significance of this structure? Which transcript is it from? Point to two specific features that help you identify its function.



d. (1 pt.) Adding arginine to the bacteria increases transcription of ORF4. What does this suggest to you about the function of the ORF4 protein? In particular, is ORF4 likely to be involved in making arginine? Explain.

3. (2 pts.) Consider a DNA sequence upstream of a gene. For simplicity, let's just look at one strand, and say that the "base sequence" is abcdefghijklmnopqrstuvwxyz. You want to explore the importance of this region in gene expression, so you make many mutants in this region. In one set of experiments, you make small deletions (say, 2-3 bases at a time). These experiments show that deletions of any bases from d to w reduce gene expression. In another set of experiments, you do extensive linker scanning tests over this region. The linker scanning results show that only changes in bases d-i and r-w reduce gene expression. Suggest a reasonable model for what this region does, based on these two sets of experiments. In particular, explain why the two experiments seem to point to different parts of the DNA as being important.

4. (6 pts.; 2 pts. per part) (continues on next page) The following is an abstract of a recent paper.

The *Escherichia coli* *proP* P2 promoter, which directs the expression of an integral membrane transporter of proline, glycine betaine, and other osmoprotecting compounds, is induced upon entry into stationary phase to protect cells from osmotic shock. Transcription from the P2 promoter is completely dependent on RpoS (σ^{38}) and Fis. Fis activates transcription by binding to a site centered at -41, which overlaps the promoter, where it makes a specific contact with the C-terminal domain of the α subunit of RNA polymerase (α -CTD). We show here that Fis and cyclic AMP (cAMP) receptor protein (CRP)-cAMP collaborate to activate transcription synergistically in vitro. Coactivation both in vivo and in vitro is dependent on CRP binding to a site centered at -121.5, but CRP without Fis provides little activation. The contribution by CRP requires the correct helical phasing of the CRP site and a functional activation region 1 on CRP. We provide evidence that coactivation is achieved by Fis and CRP independently contacting each of the two α -CTDs. Efficient transcription in vitro requires that both activators must be preincubated with the DNA prior to addition of RNA polymerase. (end of abstract)

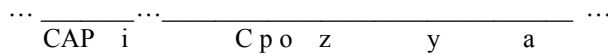
Answer the following parts (next page) based on the information in this abstract. The parts are substantially independent. (The particular σ factor discussed is of no concern for the questions below.)

a. Sketch the genome over the region discussed here. Your sketch should show the transcriptional start site, the gene whose transcription is of interest, and all the features discussed in or clearly implied by the abstract. (You need not show the genes that code for regulatory proteins; their locations are not indicated or known here.) (You can use the *lac* operon sketch on p 6 as a guide to style and level of detail needed.)

b. Using the features of this system, give an example of a genetic test that would show cis dominance. That is, describe a diploid strain for this region and describe the result that would show cis dominance. Be sure to explain what is meant by cis dominance. (You can add a reporter gene, as needed.)

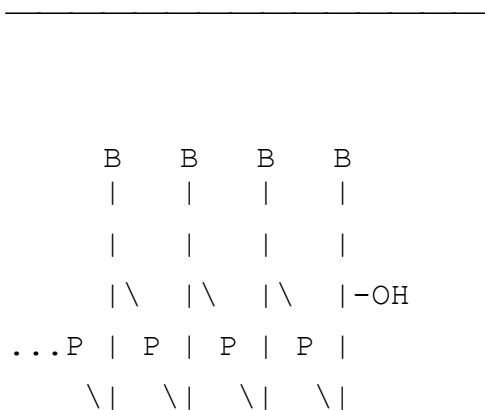
c. Sketch the “pre-initiation complex” -- the DNA with appropriate proteins bound, ready to initiate transcription. Try to incorporate as many features described in the abstract as possible. (The core RNA Pol subunit composition is $\alpha_2\beta\beta'$.)

The sketch below is for your general guidance; it shows the general (simplified) organization of the *lac* operon:



The CAP (= CRP) and i proteins are the regulatory proteins, which act at the sites C or o, respectively. p is the promoter; the remaining letters denote genes for the *lac* enzymes.

5. (3 pts.) The diagram below shows part of a growing RNA chain. Each | represents one position of the ribose sugar. (P = one phosphate group; B = any of the four normal bases. I have omitted the 2'-OH, which is not relevant to the question.)



a. (1 pt.) I showed horizontal lines above and below the main chain. Which (one) of those horizontal lines (best) represents where the template strand would be? Mark it, and label the 5' and 3' ends of this template strand.

b. (2 pts.) The immediate precursor for RNA synthesis is an activated nucleotide. Using the same diagram system for a nucleotide that I used in the main drawing, show the next nucleotide approaching the chain, about ready to form a bond to extend the chain. Show the activated structure of the incoming nucleotide and where it is about to bond. Use an arrow to show where/how the activated nucleotide precursor will connect to the chain.